

## *In Vivo* and *In Vitro* Anti-Tumor Effect of 14F7 Monoclonal Antibody

ADRIANA CARR,<sup>1</sup> CIRCE MESA,<sup>1</sup> MARÍA DEL CARMEN ARANGO,<sup>2</sup>  
ANA MARÍA VÁZQUEZ,<sup>1</sup> and LUIS ENRIQUE FERNÁNDEZ<sup>1</sup>

### ABSTRACT

The 14F7 monoclonal antibody (MAb) is an IgG<sub>1</sub> antibody that reacts specifically with GM3 (NeuGc) and with tissue sections of human tumors. We demonstrated here that this MAb is agglutinin that specifically agglutinated horse erythrocytes. Additionally, the capacity of 14F7 MAb to mediate cytotoxicity against GM3 (NeuGc)-positive murine myeloma cells, *in vitro* and *in vivo*, was evaluated. High concentrations of 14F7 MAb were needed to induce complement-dependent cytotoxicity (CDC) and antibody-dependent cell-mediated cytotoxicity (ADCC) against the murine myeloma cells. The most relevant finding was the ability of this MAb to directly kill the target cells without participation of complement. This cytotoxicity was dependent on the temperature and MAb concentration and the number of the target cells. *In vivo*, the passive treatment with 14F7 MAb produced a strong anti-tumor activity, similar to the anti-tumoral response obtained with standard chemotherapy treatment.

### INTRODUCTION

CHANGES IN THE GANGLIOSIDE EXPRESSION between tumors and normal tissues have been described.<sup>(1)</sup> In Oncology, monoclonal antibodies (MAbs) against gangliosides have gained clinical interest due to their effect in the induction of regression of cutaneous melanoma and metastases of neuroblastoma.<sup>(2-5)</sup>

Complement and cellular-dependent cytotoxicity have been reported as possible effector anti-tumor mechanisms for anti-ganglioside antibodies.<sup>(6,7)</sup> In addition, it has been proposed that the role of IgM anti-ganglioside antibodies is the abrogation of the immunosuppression induced by gangliosides shed from tumor.<sup>(8)</sup>

The *N*-glycosylated form of sialic acid (NeuGc) is not expressed in human normal tissues.<sup>(9,10)</sup> In contrast, the expression of NeuGc has been detected on tumor cells by monoclonal or polyclonal antibodies of chicken, human or murine origin.<sup>(11-13)</sup> Additionally, increased levels of NeuGc in human breast tumors were detected by biochemical methods.<sup>(14)</sup> These findings make this antigen a potential target for immunotherapy.

We recently reported the generation of a murine IgG1 MAb, named 14F7, highly specific against GM3 (NeuGc) that strongly recognized human melanoma and breast tumors.<sup>(15)</sup> In this paper, we demonstrate that 14F7 MAb is an inefficient antibody-

dependent cell-mediated cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) mediator but, this MAb has the outstanding property of causing cell death independent of complement in GM3 (NeuGc) bearing murine tumor cells. Also, we show that the treatment of tumor-bearing mice with 14F7 MAb induces a strong anti-tumor activity.

### MATERIALS AND METHODS

#### *Animals*

Balb/c, C57BL/6 and nu/nu NMRI mice, 6 to 8 weeks old, were purchased from the Center for Laboratory Animal Production (CENPALAB, Havana, Cuba). Mice were housed in plastic cages under standard conditions with access to rodent chow and water *ad libitum*. Animal care was in accordance with our institutional guidelines.

#### *MAbs*

14F7 MAb (IgG1,  $\kappa$ ) was generated by immunization of Balb/c mice with GM3 (NeuGc) ganglioside hydrophobically conjugated with human very-low-density-lipoproteins (VLDL).<sup>(15)</sup> P3 MAb

<sup>1</sup>Center of Molecular Immunology, P.O. Box 16040, Havana 11600, Cuba.

<sup>2</sup>National Radiobiology and Oncology Institute, Havana, Cuba.

TABLE 1. HEMAGGLUTINATION CAPACITY OF 14F7 MAB

Monoclonal antibody	Erythrocyte source			
	Horse	Dog	Human	Monkeys
14F7 (4°C)	+++	-	-	-
14F7 (37°C)	+++	-	-	-
P3 (4°C)	-	-	-	-
P3 (37°C)	-	-	-	-

Scored based on the MAB required for HA.  
+++ = 0.5–2 µg/mL; - = >100 µg/mL.

was used as controls. P3 MAB is (IgM, κ) that reacts with a broad battery of NueGc sialic acid containing-gangliosides.<sup>(14)</sup>

### Cells

**Tumor cell line.** The murine tumor cell lines used were P3 X63. Ag8. 653 (myeloma), B16-F10 (melanoma) and 3LLc (lung carcinoma). Cells were grown in RPMI-1640 medium (Gibco, Gaithersburg, MD), supplemented with 10% fetal calf serum (FCS), 100 U/mL penicillin and 100 µg/mL streptomycin. Prior to mice injection, B16-F10 and 3LLc cells were trypsinized and P3 X63 Ag8. 653 were resuspended by pipetting. Cells were washed and resuspended in phosphate-buffered saline (PBS), pH 7.4. Tumor cells were counted and viability was greater than 90%, as assayed by trypan blue exclusion test. Only, the myeloma cells were positive for the expression of GM3 (NeuGc) and for the recognition by 14F7 MAB, as assessed by TLC-immunostaining, immunohistochemistry, and flow cytometry analysis.

**Erythrocytes.** Horse, dog, monkey, and human erythrocytes were obtained, washed and resuspended at 2% v/v in 0.9% sodium chloride. Among them, only those of horse origin expressed GM3 (NeuGc).<sup>(16)</sup>

### Hemagglutination assay

The hemagglutination assay was performed as previously reported.<sup>(17)</sup> Briefly, various concentrations of 14F7 or P3 MABs (1–100 µg/mL) were mixed with 2% erythrocyte suspensions in round-bottomed well microplates. Then, the plate was incubated 1 h on ice or at 37°C. The end point was scored as the lowest MAB concentration that produced erythrocyte agglutination.

### In vitro cytotoxicity assays

CDC assay was performed as described.<sup>(18)</sup> Briefly, P3 X63 Ag 8. 653 myeloma cells were labeled with <sup>51</sup>Cr (Amersham, Laboratories, Buckingham, UK) for 2 h in serum-free medium at 37°C, followed by three washes. The target cells (10<sup>4</sup> cell/well) were incubated with different concentrations of 14F7 MAB (spontaneous release is measured by incubation of the target cells with medium alone). P3 MAB, which also recognizes the myeloma cells in flow cytometry experiments, was used as control antibody. The target cells (10<sup>4</sup> cell/well) were incubated with different concentrations of 14F7 MAB (10 and 50 µg/mL)

for 30 min at 37°C in the presence or absence of rabbit complement (CENPALAB, Havana, Cuba). Plates were centrifuged at 500 × g for 5 min and the supernatant (100 µL) was collected and radioactivity counted in a gamma-scintillation counter. The percentage of specific cytotoxicity was calculated as follows: % cytotoxicity = 100 × (experimental release-spontaneous release) / (maximum release-spontaneous release), where the maximum release was produced by incubation of the target cells with 10% Triton X-100 and spontaneous release is that measured by incubation of the target cells with medium alone. As control antibody was used P3 MAB, which also recognize the myeloma cells in flow cytometry experiments.

ADCC was done, as previously reported.<sup>(19)</sup> Lymphocytes harvested from spleen of BALB/C and C57BL/6 mice were used as effector cells. The murine myeloma cells were labeled with <sup>51</sup>Cr, as described above, and used as target cells. Targets were incubated with 50–100 µg/mL of 14F7 MAB for 30 min at 37°C, and washed. Labeled target cells, treated with or without 14F7 MAB, were incubated with various numbers of effector cells for 18 h at 37°C. The same control MAB used in CDC experiments was also included. The percentage of lysis was calculated as described before.

Also, the effect of 14F7 MAB on the viability of P3X63 Ag8 653 myeloma cells was studied by flow cytometry using the propidium iodide (PI) exclusion assay.<sup>(20)</sup> Different number of myeloma cells (2 × 10<sup>5</sup>–10<sup>6</sup>/100 µL) were mixed with 50 µL of different concentrations of 14F7 MAB (0.01–1 mg/mL). Cells were incubated for 30 min at 4°C and then the cells were washed with PBS, pH 7.4. Finally, cells were resuspended in 600 µL of PBS and 10 µL of PI (0.2 mg/mL) (Sigma, St. Louis, MO) was added. The analysis was performed by flow cytometry using fluorescent activated cell analyzer (Cytoron, Ortho Diagnostic System, Raritan, NJ). P3 (an IgM MAB, which recognizes the same antigen) MAB was used as control antibody.

### In vivo effect of 14F7 MAB on tumor cell growth in mice

Two different types of experiments were performed. (a) Different murine solid tumors, expressing or not GM3 (NeuGc) were used. With P3 X63 Ag8 653 myeloma cells, to obtain solid tumors, the cells were inoculated in nu/nu NMRI mice (*n* = 5) and the animals received 1 × 10<sup>6</sup> myeloma cells intramuscularly (i.m.) (Day 0). On Days 0, 2, 4, 6, 8, and 10, mice were injected intraperitoneally (i.p.) with 200 µg of 14F7 MAB in PBS. Similar experiments were performed using C57BL/6 mice (*n* = 10) inoculated with 2 × 10<sup>3</sup> B16-F10 cells or 5 × 10<sup>4</sup>

TABLE 2. COMPLEMENT-DEPENDENT CYTOTOXICITY MEDIATED BY 14F7 MAB

MAB (µg/mL)		% of lysis	
		Rabbit serum dilution 1:5	1:10
14F7	50	51	45
	10	4	2
P3	50	65	65
	10	62	61

TABLE 3. CYTOTOXICITY INDUCED WITHOUT COMPLEMENT

MAb (mg/mL)	Target cells	Cell concentration		
		10 <sup>7</sup> cells/mL (%)	0.5 × 10 <sup>7</sup> cells/mL (%)	10 <sup>6</sup> cells/mL (%)
P3 (0.1)	P3 × 63 Ag8 653 myeloma	5.9 <sup>a</sup>	ND	ND
P3 (0.01)	P3 × 63 Ag8 653 myeloma	4.5	ND	ND
14F7 (1)	P3 × 63 Ag8 653 myeloma	80	36	5%
14F7 (0.4)	P3 × 63 Ag8 653 myeloma	63	ND	ND
14F7 (0.1)	P3 × 63 Ag8 653 myeloma	62	10	6%
14F7 (0.01)	P3 × 63 Ag8 653 myeloma	7	6	5.9%

<sup>a</sup>Percent of dead cells measured using the PI exclusion assay. ND: not determined.

3LLc. Also, a group of nu/nu NMRI mice were inoculated with the myeloma cells, but the treatment with 14F7 MAb started on Day 10.

(b) Balb/c mice (*n* = 10) were injected i.p. with 0.5 mL of incomplete Freund's adjuvant. After 3 days, animals were inoculated ip with 10<sup>4</sup> myeloma (Day 0). Animals were treated i.v. on Days 0, 2, 4, 6, 8, and 10 with 100 or 200 µg of 14F7 MAb. Animals that were injected iv with 20 mg/kg of cyclophosphamide (Lemery S.A., Mexico, D.F., Mexico) weekly were used as the positive control group.<sup>(21)</sup> Mice that received PBS instead of the MAb were used as negative controls.

*Statistical analysis.* The two largest perpendicular diameters of tumors were recorded with a caliper, twice a week to evaluate tumor growth. Analysis of variance (ANOVA) test was used to compare mean tumor volumes among each group. Survival was analyzed by Kaplan–Mayer method, and groups were compared by log–rank test, *p* < 0.05 was considered as significant and *p* < 0.01 as highly significant.

**RESULTS**

The specific hemagglutination capacity of 14F7 MAb is shown in Table 1. 14F7 MAb was able to agglutinate horse erythrocytes, but not those from dog, monkey, or human origin. The difference among these different types of erythrocytes is that in horse erythrocytes, 95% of the lipid containing sialic acid is in the form of GM3 (NeuGc)<sup>(16)</sup>; while this ganglioside is absent in the other erythrocyte species studied. The agglutination was observed both at 37°C and at 4°C. Although, P3

MAb recognizes GM3 (NeuGc), it was not able to agglutinate horse erythrocytes.

14F7 MAb incubated with myeloma cells in the presence of rabbit complement was tested for its ability to lyse these GM3 (NeuGc)-positive cells. Table 2 shows the results of this experiment. The incubation for 30 min at 37°C with 50 µg/mL of 14F7 produced 50 and 45% cytotoxicity of the target cells, depending on the rabbit complement dilution used. Lower concentration of 14F7 MAb (10 µg/mL) did not produce cytotoxicity. P3 MAb was able to produce more than 60% of lysis at both concentrations.

To assess whether 14F7 could induce ADCC, P3 X63 Ag8 653 myeloma cells were labeled with <sup>51</sup>Cr and incubated in the presence of the MAb and the effector cells. 14F7 MAb induced lysis of target cells equal or higher than 30% only when the ratio of mouse splenocytes:target cells was 80:1 and when MAb was used at a concentration of 100 µg/mL (Table 3). The control P3 MAb produced similar cytotoxicity effect than 14F7 MAb. No cytolytic effect was detected when target cells were incubated only in the presence of the antibody. The percent of <sup>51</sup>Cr release by effector cells alone (without the antibody), was always between 1–10%.

In the propidium iodine cytotoxicity exclusion assay performed at 4°C for 30 min, the percent of myeloma cells in the PI+ population (dead cells) correlated directly to 14F7 MAb and target cell concentrations, and not with P3 MAb concentration. The best cytotoxicity with 14F7 MAb was induced at the concentration of 10<sup>7</sup> myeloma cells/mL (Table 4).

The role of 14F7 MAb in modulating the growth of GM3 (NeuGc)-positive and -negative tumors was studied (Fig. 1). In the experiment nu/nu NMRI mice were inoculated on day 0

TABLE 4. ANTIBODY-DEPENDENT CELLULAR CYTOTOXICITY MEDIATED BY 14F7 MAb AGAINST MURINE MYELOMA CELLS

MAb	µg/mL	(% of Lysis C57BL6 splenocytes E/T ratio <sup>a</sup> )			(% of Lysis Balb/c splenocytes E/T ratio)		
		20:1	40:1	80:1	20:1	40:1	80:1
14F7	50	12.6	24.1	26.7	8.3	27.8	36.8
14F7	100	7.5	26.6	42.1	9.6	28.3	44.8
P3	50	8.8	11.5	21.5	8.9	22.2	39.4
P3	100	4.5	22	38	11	25.2	39

<sup>a</sup>E/T ratio: effector:target cells ratio.

with  $1 \times 10^6$  murine myeloma cells and were treated iv with 14F7 MAb or with PBS on Days 0, 2, 4, 6, 8, and 10. A significant suppression of tumor growth was observed in the mice that received passive therapy with 14F7 MAb, in comparison with the control group ( $p < 0.01$ ). In contrast, when the treatment with the MAb started 10 days after the inoculation of the myeloma cells, no differences were observed when compared with the control mice (Fig. 1B). No differences were detected in tumor growth between 14F7 MAb-treated and PBS-treated C57BL/6 mice inoculated with B16-F10 or 3LLc tumor cells (antigen negative) (Figs. 1C and D).

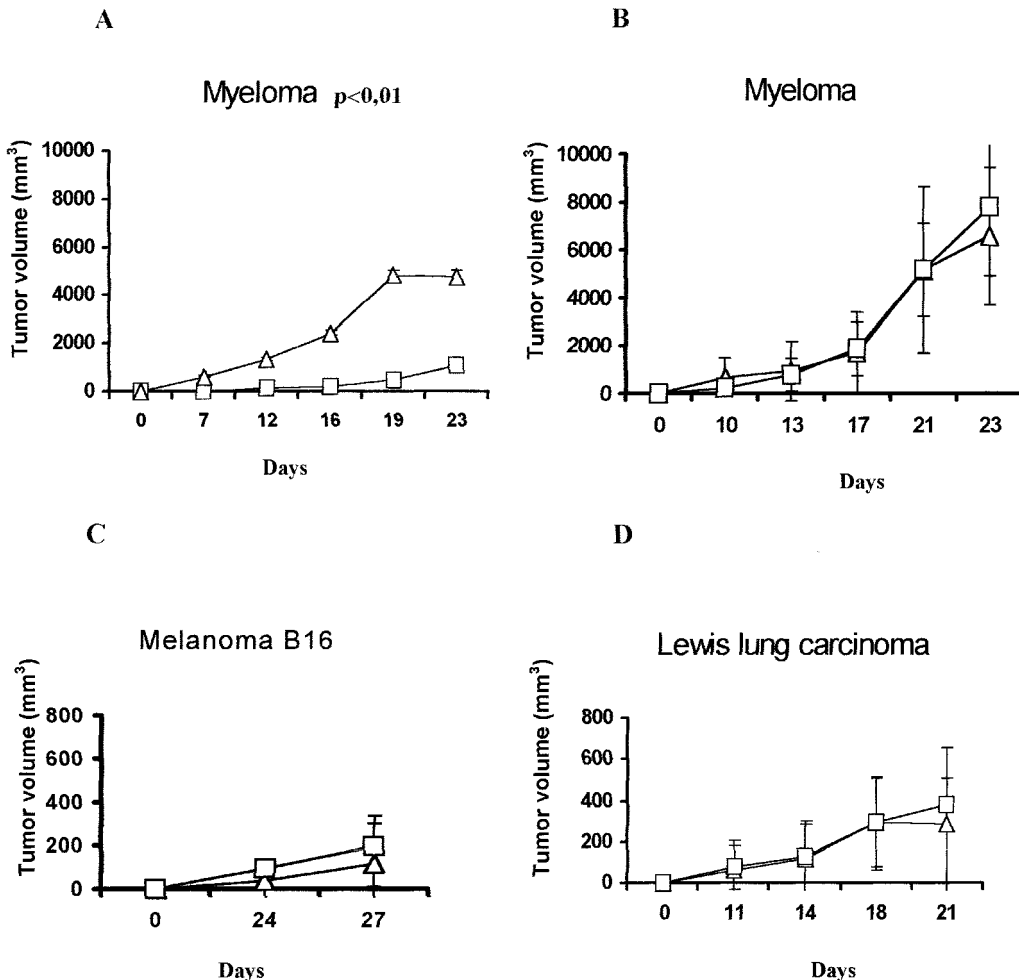
Balb/c mice bearing P3 X63 Ag8 653 ascitic tumors were kept until moribund in order to study the effect of the passive treatment with 14F7 MAb on animal survival, in comparison with the effect produced by chemotherapy treatment with cyclophosphamide as positive control. Figure 2 shows the results of this experiment. The survival of tumor-bearing animals was significantly prolonged in the groups of animals treated i.v. with 100  $\mu\text{g}$  ( $p = 0.007$ ) or 200  $\mu\text{g}$  ( $p = 0.003$ ) of 14F7 MAb, in comparison with the group that received only PBS. On the other

hand, no differences in the prolongation of survival were observed among the groups treated with 14F7 MAb and the ones treated with cyclophosphamide ( $p = 0.05$ ).

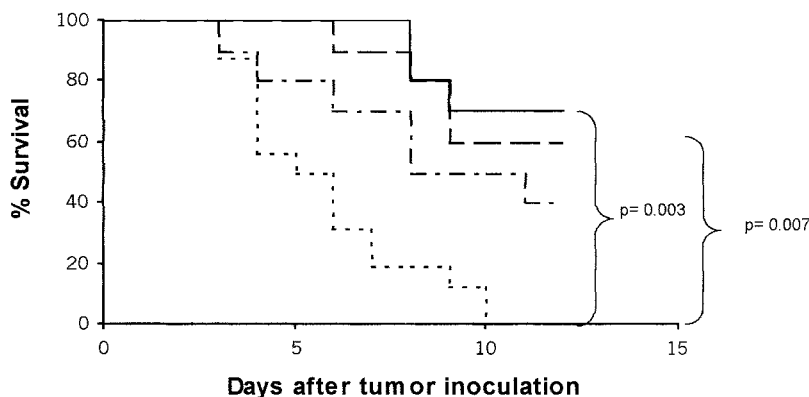
## DISCUSSION

In a previous study, we described that 14F7 MAb binds specifically to GM3 (NeuGc), and that it also reacts with human breast and melanoma tumors; in contrast, with its low reactivity in normal tissues.<sup>(15)</sup> In this study, the anti-tumor activity of 14F7 MAb against murine tumor cells expressing GM3 (NeuGc) was observed and the possible effector mechanisms through which this antibody exert its biological effect was analyzed.

CDC and ADCC are effector mechanisms that have been suggested to be involved in the antibody effect against some tumors expressing gangliosides.<sup>(22,23)</sup> Nevertheless, with 14F7 MAb high concentrations of the antibody are needed to induce



**FIG. 1.** Effect of 14F7 MAb on murine tumor growth. Mice were treated with 14F7 MAb or with PBS (control) (A), (C), and (D) represent the tumor growth in mice treated with 14F7 MAb on days 0, 2, 4, 6, 8 and 10. (B) shows the results obtained when the treatment with 14F7 MAb started on Day 10. Data represent means  $\pm$  SEM.  $p < 0.01$ , ANOVA test.



**FIG. 2.** Survival of 14F7-treated mice. Treatments were as follows: ———100  $\mu\text{g}/\text{mL}$  of 14F7 MAb, ——— 200  $\mu\text{g}/\text{mL}$  of 14F7 MAb, ——— 200  $\mu\text{g}$  of cyclophosphamide and - - - - - PBS. Comparison of overall survival between the different groups was tested by Log-rank test.

cytotoxic effects by both mechanisms against myeloma cells, in spite of their high expression of GM3 (NeuGc). Thus, 14F7 MAb is an inefficient ADCC and CDC mediator. This result is in agreement with the fact that murine IgG<sub>1</sub> antibodies are weak inducers of CDC and ADCC.<sup>(24)</sup>

On the other hand, 14F7 MAb showed a strong anti-tumor activity against the myeloma cells *in vivo*. The weak CDC and ADCC activity could be involved, but another possible explanation for this phenomenon is the characteristic of this MAb to produce a complement independent cytotoxicity at low temperature. Apoptosis does not seem to be involved in the loss of cellular viability since time to cell death is short not corresponding with the duration of events in the programmed cell death mechanism. This uncommon property was previously described for other MABs. One of them, named RE2, was found while searching for a xenogeneic antibody specific for mouse T-cell membrane molecules. This antibody at 4°C produced a complement-independent lysis of a T-cell clone used for MABs screening.<sup>(25)</sup> The other examples were human VH4-34 gene-derived anti-i cold agglutinin (CA) MABs that bound human and murine B cells.<sup>(26)</sup> The binding of B cells by these MABs led also to a rapid noncomplement dependent cytotoxicity at 4°C. Similar to these MABs, the cytotoxicity produced by the incubation of target cells with 14F7 MAB is dependent on the temperature and MAB concentration, but not on complement activity.

The mechanisms by which 14F7 induces cell death are still unknown, but as was proposed for RE2,<sup>(26)</sup> a cross-linking of GM3(NeuGc) molecules on the surface of target cells by 14F7 MAB might induce complement-independent cell lysis.

It is noteworthy that P3 MAB, an IgM antibody that also recognizes GM3(NeuGc), was incapable of agglutinating horse erythrocytes. While, 14F7 MAB, an IgG<sub>1</sub> antibody, was able to agglutinate these erythrocytes at 37°C and at 4°C. Interestingly, the VH4-34 antibody is a cold agglutinin that also induces complement independent cell death.<sup>(26)</sup>

Growth inhibition and prolonged survival of the myeloma tumor are evidences of anti tumor effect obtained after treatment with 14F7 MAB. This result correlated with the high ex-

pression of GM3 (NeuGc) ganglioside as target molecule, shown by biochemical and immunohistochemistry analysis in this murine myeloma cell line. Furthermore, no anti-tumor effects were observed with the antigen negative cell lines, the antigen negative murine tumors LLC and B16.

In this study we demonstrate that when nu/nu NMRI mice were treated ip with 14F7 MAB the same day of myeloma cell inoculation, a suppression of tumor growth was observed; but when the MAB treatment started 10 days after challenge, no anti-tumor effect was observed. This result indicates that the use of 14F7 MAB could be effective mainly in an adjuvant therapy, as has been proposed for other MABs that react with other tumor-associated antigens.<sup>(4)</sup>

Therapy with cyclophosphamide produces tumor regression in murine myelomas.<sup>(21)</sup> Our results show that treatment with 14F7 MAB produced an increase in anti-tumor effect related to MAB dose similar to the cyclophosphamide chemotherapy. Nevertheless, optimization of the administration frequency and MAB concentration should be performed.

New clinical evidences of anti tumor effect of MABs in cancer patients have appeared in the last years.<sup>(27,28)</sup> Most MABs developed for treatment of solid tumors are indicated combined with chemotherapy or radiotherapy. The evidences of anti-tumor effect shown here with 14F7 MAB in experimental solid tumors indicate its potential use for passive immunotherapy in these tumor types.

A phase I tumor imaging clinical trial with 14F7 MAB in advanced cancer patients will be carried out in the near future in order to define the capacity of this MAB to target tumors *in vivo*, as a critical result to use it in passive immunotherapy.

## ACKNOWLEDGMENTS

The authors thank Dr. Franz Torres for statistical advice, technician Dariel Morales for valuable support and supervision at the animal care facility; and Dr. Blanca Rosa Tormo for their helpful advice and for editorial assistance. Financial support

was provided in part by Laboratorios ELEA (Buenos Aires, Argentina) and Cuba Government.

## REFERENCES

- Hakomori S.H: Tumor associated carbohydrate antigens. *Ann Rev Immunol* 1984;2:103-126.
- Houghton AN, Mintzer D, Cordon-Cardo C, Welt B, Fliegel S, Vadhan S, Carswell E, Melaned MR, Oettgen, HF, and Old LJ: Mouse monoclonal antibody IgG3 antibody detecting GD3 ganglioside. A phase I trial in patients with malignant melanoma. *Proc Natl Acad Sci USA* 1985;82:1242-1246.
- Irie R, and Morton DL: Regression of cutaneous metastatic melanoma by intralesional injection with human monoclonal antibody to ganglioside GD2. *Proc Natl Acad Sci USA* 1986;83:8694-8698.
- Zhang H, Zhang S, Nai-kong V, Cheung V, Ragupathi G, and Livingston PO: Antibodies against GD2 ganglioside can eradicate syngenic cancer micrometastases. *Cancer Res* 1998;58:2844-2049.
- Kushner BH, Kramer K, and Cheung NK: Phase II trial of the anti-GD2 monoclonal antibody 3F8 and granulocyte-macrophage colony-stimulating factor for neuroblastoma. *J Clin Oncol* 2001; 19:4189-4194.
- Mujoo K, Kipps TJ, Yang HM, Cheresch DA, Wargalla U, Sander DJ, and Reisfeld RA: Functional properties and effect on growth suppression of human neuroblastoma tumors by isotype switch variants of monoclonal anti-ganglioside GD2 antibody 14.18. *Cancer Res* 1989;49:2857-2861.
- Livingston PO, and Ragupathi G. Carbohydrate vaccines that induce antibodies against cancer. 2. Previous experience and future plans. *Cancer Immunol Immunother* 1997;45:10-19.
- Ravindranath MH, Gonz ales AM, Nishimoto K, Tam WY, Sah D, and Morton DL: Immunology of gangliosides. *Indian J Exp Biol* 2000;38:301-312.
- Leeden RW, and Yu RK: Chemistry and analysis of sialic acid. In: Rosemberg A and Schegtrund CL (Eds.). *Biological Role of Sialic Acid*. New York, Plenum Press, 1976, pp. 1-48.
- Kawai T, Kato A, Higashi H, Kato S, and Naiki M: Quantitative determination of N-glycolyl-neuraminic acid expression in human cancerous tissues and avian lymphoma cell lines as a tumor associated sialic acid by gas chromatography-mass spectrometry. *Cancer Res* 1991;51:1242-1246.
- Koda T, Shimosakoda T, Asaoka H, Nishinaka S, Tamura I, Nakaba H, and Matsuda H: Detection of Hanganutziu-Deicher antigen in patients with hepatocellular carcinoma. *Intern Hepatol Commun* 1994;2:310-315.
- Furukawa K, Yamaguchi H, Oettgen, HF, Old, LJ, Lloyd, and KO: Analysis of the expression of N-Glycolylneuraminic Acid-containing Gangliosides in Cell and Tissues using Two Human Monoclonal Antibodies. *J Biol Chem* 1988;263:18507-18512.
- V azquez AM, Alfonso M, Karlsson A, Carr A, Barroso O, Fern andez LE, Rengifo E, Lanio ME, Alvarez, C, Zeuthen J, and P erez R: Generation of a murine monoclonal antibody specific for N-glycolylneuraminic acid containing gangliosides that also recognizes sulfated glycolipids. *Hybrid* 1995;14:551-556.
- Marquina G, Waki H, Fern andez LE, Kon K, Carr A, Valiente O, P erez R, and Ando S: Gangliosides Expressed in Human Breast Cancer. *Cancer Res* 1996;56:5165-5171.
- Carr A, Mullet A, Mazorra Z, V azquez, AM, Alfonso M, Mesa C, Rengifo E, P erez R, and Fern andez LE: Mouse IgG<sub>1</sub> monoclonal antibody specific for N-glycolyl GM3 ganglioside recognized breast and melanoma tumors. *Hybridoma* 2000;19:241-247.
- Higashi H, Naiki M, Matuo S, and Okouchi K: Antigen of "serum sickness" type Heterophile antibodies in pathologic human sera: identification as gangliosides with N-Glycolylneuraminic acid. *Biochem Biophys Res Commun* 1977;79:388-395.
- Kasukawa R, Kano K, Bloom ML, and Milgrom F: Heterophile antibodies in pathologic human sera resembling antibodies stimulated by foreign species sera. *Clin Exp Immunol* 1976;25:122-132.
- Herlyn DM, and Koprowsky H. Monoclonal anticolon carcinoma antibodies in complement-dependent cytotoxicity. *Int J Cancer* 1981;26:769-774.
- Schmid I, Uttenbogaart CH, and Giorgy JV: Sensitive method for measuring apoptosis and cell surface phenotype in human thymocytes by flow cytometry. *Cytometry* 1994;15:12-20.
- Dohi T, Nores G, and Hakomori S. An IgG<sub>3</sub> monoclonal antibody established after immunization with GM3 lactone: immunochemical specificity and inhibition of melanoma cell growth in vitro and in vivo. *Cancer Res* 1988;48:5680-5685.
- Broun Go JR, Petruska PJ, Hiramoto RN, and Cohen HJ: Cisplatin, BCNU, cyclophosphamide, and prednisone in multiple myeloma. *Cancer Treat Rep* 1982;66:237-242.
- Chapman PB, Lonberg M, and Houghton AN: Light chain variants of an IgG3 anti-GD3 monoclonal antibody and the relationship among avidity, effector functions, tumor targeting, and antitumor activity. *Cancer Res* 1990;50:1503-1509.
- Cheung NK, Lazarus H, Miraldi FD, Abramowsky CR, Kallick S, Saarinen UM, Spitzer T, Strandjord SE, Coccia PF, and Berger NA: Ganglioside GD2 specific monoclonal antibody 3F8: a phase I study in patients with neuroblastoma and malignant melanoma. *J Clin Oncol* 1987;5:1430-1440.
- Rosales C, Jeglum KA, Obrocka M, and Steplewski Z: Cytolytic activity of murine anti-dog lymphoma monoclonal antibodies with canine effector cells and complement. *Cell Immunol* 1988;115: 420-428 .
- Matsuoka S, Asano Y, Sano K, Kishimoto H, Yamashita I, Yorifuji H, Utsuyama M, Hirokawa K, and Tada T: A novel type of Cell Death of Lymphocytes Induced by a Monoclonal Antibody without Participation of complement. *J Exp Med* 1995;181: 2007-2015.
- Bhat NM, Bieber MM, and Teng NNH: Cytotoxicity of murine B lymphocytes induce by human VH4-34(VH4.21) gene-encoded monoclonal antibodies. *Clin. Immunol. Immunopath* 1997;84:283-289.
- Robert F, Ezekiel MP, Spencer SA, Meredith RF, Bonner JA, Khazaeli MB, Saleh MN, Carey D, Lobuglio AF, Wheeler RH, Cooper MR, and Waksal HW: Phase I study of anti-epidermal growth factor receptor antibody cetuximab in combination with radiation therapy in patients with advanced head and neck cancer. *J Clin Oncol* 2001;19:3234-3243.
- Livingston RB, and Esteva FJ: Chemotherapy and herceptin for HER2(+) metastatic breast cancer: the best drug? *Oncologist*, 2001;6:315-316.

Address reprint requests to:  
*Adriana Carr, Ph.D.*  
*Center of Molecular Immunology*  
*P.O. Box 16040*  
*Havana 11600*  
*Cuba*

*E-mail: adriana@ict.cim.sld.cu*

Received for publication September 4, 2002. Accepted for publication September 20, 2002.